

# *S. aureus* Gyrase Microplate Assay Kit



## Product Description (Product Number # SATRG01, SATRG02)

The kit is supplied with sufficient *S. aureus* gyrase enzyme, plasmid DNA substrate (relaxed pNO1; supplied at 1 mg/ml), 5X Assay Buffer, Enzyme Dilution Buffer, DNA-detection dye (Promega Diamond dye) and TFO1 oligo for 100 assays. The enzyme is supplied at a concentration of 10 U/ $\mu$ l in Dilution Buffer (see Note 1). The kit is also supplied with sufficient Wash Buffer, TF buffer and T10 buffer for one 96-well plate. These buffers are supplied as 20X concentrates and must be diluted prior to use with ultra-pure water.

Store at -80 °C. (Stable for 3 months undiluted.) The enzyme should be centrifuged briefly before opening and it is recommended that it is aliquoted (undiluted) to avoid repeated freeze-thaw cycles.

**For *in vitro* laboratory research use only.**

### Dilution Buffer

50 mM Tris.HCl (pH 7.5)  
1 mM DTT  
1 mM EDTA  
40 % (w/v) glycerol

### Assay Buffer (supplied as 5X stock)

40 mM HEPES.KOH (pH 7.6)  
10 mM magnesium acetate  
10 mM DTT  
2 mM ATP  
500 mM potassium glutamate  
0.05 mg/ml albumin

### TF Buffer (supplied as a 20X stock)

50 mM sodium acetate (pH 5.0)  
50 mM NaCl  
50 mM MgCl<sub>2</sub>

### Wash Buffer (supplied as a 20X stock)

20 mM Tris.HCl (pH 7.6)  
137 mM NaCl  
0.005 % (w/v) BSA  
0.05 % (v/v) Tween-20

### DNA-detection Dye (supplied as a 1,000X stock)

Supplied in 100% DMSO

### T10 Buffer (supplied as a 20X stock)

10 mM Tris.HCl (pH 8)  
1 mM EDTA

## Preparation of Plate and Supercoiling Assay

Rehydrate wells with 3 x 200  $\mu$ l Wash Buffer (diluted from 20X stock before use).

Immobilize 100  $\mu$ l of 500nM TFO1 oligo in each well (5  $\mu$ l of 10  $\mu$ M TFO1 in 95  $\mu$ l Wash Buffer), 5 minutes at room temperature. Wash off excess oligo with 3 x 200  $\mu$ l Wash Buffer.

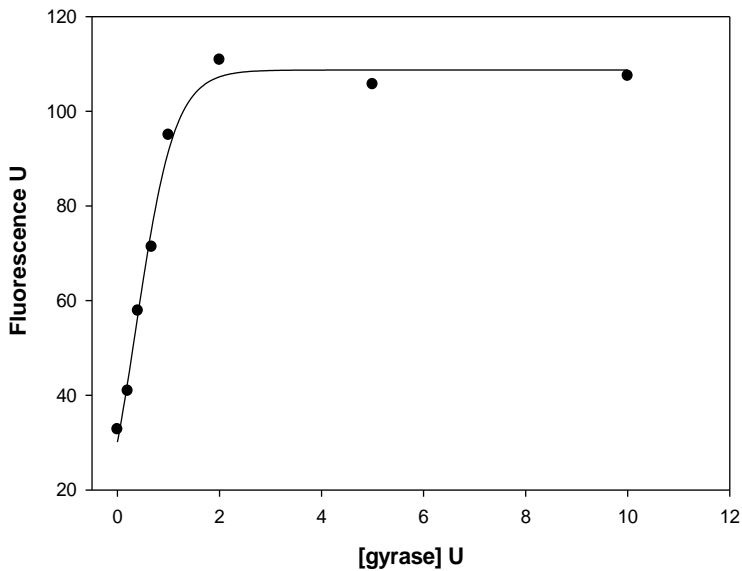
Incubate 1.5 U of *S. aureus* gyrase with 0.75  $\mu$ g of relaxed pNO1 in a reaction volume of 30  $\mu$ l at 37°C for 30 minutes in Assay Buffer. Incubate reaction in well.

Add 100  $\mu$ l TF Buffer (diluted from 20X stock before use) to well and incubate for a further 30 minutes at room temperature to allow triplex formation.

Remove liquid from well and wash with 3 x 200  $\mu$ l TF Buffer to remove unbound plasmid.

Stain with DNA-detection dye (diluted to 1X with T10 buffer. Add 200  $\mu$ l per well. Incubate for 10 - 20 minutes, **mix** and read in fluorescence plate reader; Ex: 495 nm; Em: 537 nm)..

**Supercoiling of relaxed pNO1 by varying amounts of *S. aureus* gyrase in the plate assay**



### Quality Control

- 1) Purity: *S. aureus* gyrase is purified to > 95 % purity as judged by SDS-polyacrylamide gel electrophoresis.
- 2) pBR322 was also incubated for 4 hrs in assay buffer + 10 mM MgCl<sub>2</sub> at 37 °C. These tests were negative for the formation of linear products, indicating the absence of nuclease contamination.
- (3) No activity was detectable when the single subunits were assayed alone.

### References

Maxwell, A., Burton, N.P. and O'Hagan, N. (2006) High-throughput assays for DNA gyrase and other topoisomerases. *Nucleic Acid Res.* **34(15)**, e104

### Notes.

1. A unit of enzyme activity is defined as the amount of enzyme that just supercoils 0.5 µg of relaxed pBR322 in 30 minutes at 37°C as judged by agarose gel electrophoresis.
2. It is important to mix the samples before reading or they will be greatly reduced.

**Patent for assay held by Inspiralis Ltd., Norwich, Norfolk, UK. (Patent No. GB0424953.8).  
Kit issued with limited licence for individual use only.**

**DNA-detection stain is Diamond nucleic acid dye supplied under license from Promega Corporation.**